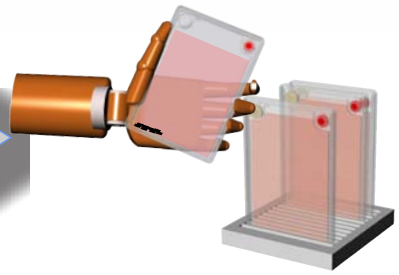
Celartia

HARVESTING and SEEDING CELLS FROM PETAKA-ET

Petaka™

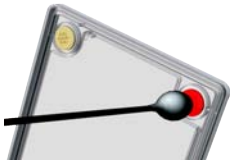
PROTOCOL: *HARVESTING and SEEDING CELLS FROM PETAKA-ET*

Take incubated Petaka ET from the incubator



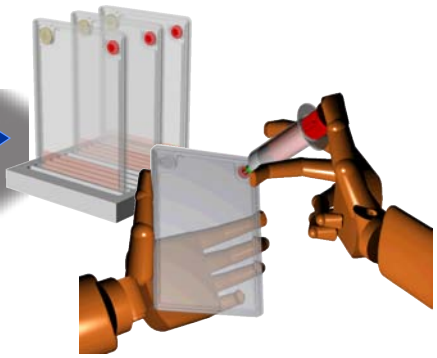
Put incubated **Petaka ET** on top of the powered Cell Flipper platform and allow the gliders to oscillate randomly for 3 to 4 min. **1**

2 Disinfect port with 90% Ethanol



Slowly withdraw all media (cell suspension) **3**

4 Transfer the cell suspension into new Petakas.



Harvesting & sub-culturing anchored cells

ADHERENT CELLS WITH PetakaG3-ET WITHOUT ENZYMES

High Cell preservation
Very high Surface protein preservation

1. Place incubated Petaka ET[1] on top of the powered Cell Flipper platform and allow the gliders to travel randomly for 3 to 4 minutes.
2. Disinfect port with 90% Ethanol
3. Slowly withdraw all media
4. Transfer the cell suspension into new Petakas.

Expanding from 1 culture to 10 subcultures, 5-6 min.

1) Petaka G3 with wipers

ADHERENT CELLS WITH TRYPSIN-EDTA FAST PROCEDURE WITH ENZYMES

High Cell preservation
Low Surface protein preservation

1. **Disinfect the port with 90% Ethanol & a flame**
2. **Completely extract all the media from the Petaka**
3. **Inject 4 ml of 0.25% Trypsin-EDTA solution**
4. **Shake gently whilst maintaining in a horizontal position until all internal surfaces are bathed in the trypsin solution**
5. **Incubate at 37ºC for 3 min in a horizontal position (on both sides)**
6. **Check Cell detachment under the microscope**
7. **WHEN CELLS ARE DETACHED! Tap the Petaka 2-3 times**
8. **Disinfect port with 90% Ethanol**
9. **Inject 2 ml Heat Inactivated FBS**
10. **Slowly withdraw all media**
11. **Transfer the cell suspension into new Petakas, and complement the media in each one.**

Expanding from 1 culture to 10 subcultures, 10-12 min.

Harvesting & sub-culturing anchored cells

ADHERENT CELLS WITH Petaka-ET WITHOUT ENZYMES OPTIMUM PROCEDURE

High Cell preservation
Very high Surface protein preservation

1. Take incubated Petaka ET from the incubator
2. Place incubated Petaka ET on top of the powered ClearCell platform and allow the wipers to travel randomly for 3 to 4 minutes.
3. Disinfect port with 90% Ethanol
4. Slowly withdraw all media
5. Transfer the single cell suspension into new Petakas.

Expanding from 1 culture to 10 subcultures, 5-6 min.

ADHERENT CELLS WITH TRYPSIN-EDTA CLASSIC PROTOCOL

Very high Cell preservation
Low Surface protein preservation

1. Disinfect the port with 90% Ethanol & a flame
2. Completely extract all the media from the Petaka
3. Inject 4 ml of 0.25% Trypsin-EDTA solution
4. Shake gently whilst maintaining in a horizontal position until all internal surfaces are bathed in the trypsin solution
5. Incubate at 37ºC for 3 min in a horizontal position (on both sides)
6. Check Cell detachment under the microscope
7. WHEN CELLS ARE DETACHED! Tap the Petaka 2-3 times
8. Pellet the cells in the bottom left hand corner using a Petaka centrifuge (see centrifugation). To pellet the cells in the bottom left hand corner of Petaka, position it with the filter being the upper most corner of the device
9. After centrifuge, the cell pellet will be visible in the bottom left hand corner
10. Disinfect port with 90% Ethanol
11. Carefully withdraw the media avoiding disruption of the cell pellet
12. Inject 10 ml of media with 10% FBS
13. Resuspend the cell pellet shaking carefully
14. Pellet the cells in the bottom right hand corner using a Petaka centrifuge (see centrifugation). To pellet the cells in the bottom right hand corner of Petaka, position it with the port being the upper most corner of the device
15. After centrifuge, the cell pellet will be visible in the bottom right hand corner
16. Disinfect port with 90% Ethanol
17. To extract the pellet, use a micro pipette with a 2mL tip
18. Transfer the cell suspension into new Petakas, and complement the media in each one.

Expanding from 1 culture to 10 subcultures, 15-20 min.